THE EFFECT OF TURMERIC AND GARLIC PHYTOBIOTIC ADDITION WITH DIFFERENT DURATION STORAGE ON THE FUNGAL COLONY IN FEED

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ABSTRACT

Turmeric and garlic phytoibiotic contain bioactive substances, each of which has a function as an antifungal. The study aims to determine the ability of turmeric and garlic phytoibiotic pressed fungal colonies in the feed with different storage times. The data were analyzed by a factorial experimental design with 5 treatments, 4 replications, and 2 repetitions. The first factor types phytoibiotic, and the second factor was storage duration. R0 = basal ration (without phytoibiotic), R1 = basal ration + 0.3% synthetic antifungal, R2 = basal ration + 2.5% turmeric, R3 = basal ration + 5% garlic, R4 = (basal ration + 2.5% turmeric + 5% garlic. W0, W1, W2, and W3 respectively with storage time 0, 2, 4, and 6 weeks. Samples were taken from each experimental unit homogeneously. The results showed the average fungal colonies of garlic phytobiotic treatment (26.06±44.76 CFU*x10^3/g) in 2-week storage time (25.20±22.52 CFU*x10^3/g) lower than the provision of synthetic preservatives (57.75±52.03 CFU*x10^3/g) and the control treatment (74.87±70.69 CFU*x10^3/g). Fungal colonies increase rapidly with increasing length of time of storage. The conclusion of this study was that the addition of turmeric and garlic phytoibiotic capable as an antifungal in feed.

Keywords: fungal colony, garlic, phytoibiotic, storage, turmeric

INTRODUCTION

Mycotoxins produced by fungi during storage aerobic and anaerobic often not detected in the ration. This has an impact on the decline in the nutritional quality of their diets. The mechanism of action of preservatives is to decrease the pH of the feed so that the fungus can not grow. Alternative materials are safe and do not leave residue was phytoibiotic use that has a function as an antifungal.

Utilization phytoibiotic as natural growth promoters (NGPs) has been identified as an effective alternative to antibiotics. Phytoibiotic as NGPs develop as a feed additive, which increases immunity, production performance and is very effective in improving the health of the digestive tract (Panda et al., 2009), stimulates livestock nutrition, antimicrobial, coccidiostatic and antihelmintic (Panda et al., 2006).

Biological response to garlic among others, include a reduced risk of cancer and antitumor, stimulation of immune function (Balasenthil et al., 2001; Song and Milner, 2001; Galeone et al., 2006; Corzo-Martinez et al., 2007). The antibacterial alisin reported to be effective against a large number of Gram positive and Gram negative including Salmonella, Staphylococcus, Streptococcus, Klebsiela, Proteus, Bacillus, Clostridium, Coliform and Lactobacillus, as well as some anaerobic bacteria (Rahman, 2007; Bongiorno, 2008; Saravanan et al., 2010).

MATERIAL and METHODS

Collection and making of turmeric and garlic powder

Turmeric Turina-2 obtained from Balai Tanaman Obat Aromatik, Bogor, West Java of Indonesia, with 10-month maturity, garlic kathing of traditional markets. Making the turmeric powder and garlic are two herbs
are washed and then thinly sliced. Turmeric and garlic slices dried at a temperature of 50°C for 3 days. Turmeric and garlic crushed in order to obtain turmeric and garlic powder.

Ration storage with the addition of phytobiotic

Turmeric and garlic powder mixed into the ration according to treatment. Many rations each experimental unit as much as 1 kg. Ration is placed on the board with a height of 10 cm from the floor. Temperature and humidity of the room is measured at any time.

Sample analysis procedure

Preparation and homogenization. Dilution of samples, take 1 gram of sample enter into NaCl solution 9 ml then pipette 1 ml of each dilution into a petri dish in simplo and Duplo. PDA (Potato Dextrose Agar) medium was added chlorampenicol 100 mg/ml after the temperature 50°C. Pouring PDA which has previously been melted and added chlorampenicol (temperature 45-50°C) 15-20 ml into the petri dish. Wiggle the petri dish such that the mixture evenly. Allow to freeze and then incubation for 3-5 days at a temperature 22-25°C or room temperature. Let stand until the colony can be calculated. Fungal colonies counted after incubation the number of colonies units/gram. Fungal colony counting with the following formula

\[ N = \frac{\Sigma C}{(1 \times n1) + (0,1 \times n2) + (0,01 \times n3)} \times 10^{-3} \]

Description:

- \( N \) = the number of colonies of the product, expressed in colony/ml or colonies/g;
- \( \Sigma C \) = number of colonies on all the bowls were calculated;
- \( n1 \) = number of bowls on the first dilution is calculated;
- \( n2 \) = number of bowls on the calculated dilution;
- \( n3 \) = number of bowls on the third dilution is calculated;
- \( d \) = dilution first calculated.

Experimental Design and Statistical Analysis

The data were analyzed by a factorial experimental design with 5 treatments 4 replications and 2 repetitions. The first factor types phytobiotic and the second factor was storage duration. R0 = basal ration (without phytobiotic), R1 = basal ration + 0.3% synthetic antifungal, R2 = basal ration + 2.5% turmeric, R3 = basal ration + 5% garlic, R4 = (basal ration + 2.5% turmeric + 5% garlic). W0, W1, W2 and W3 respectively with storage time 0, 2, 4 and 6 weeks. The treatments means with significant differences at P<0.05 were compared using Least Significant Difference (LSD) (Gomez and Gomez, 1984).

RESULTS and DISCUSSION

The average number of fungal colonies in feed for different storage with the addition of turmeric and garlic phytobiotic can be seen in Table 1.

Table 1. Average number of fungal colonies in feed during storage contrast (CFUx10³/g)

<table>
<thead>
<tr>
<th>Treatments</th>
<th>Time (weeks)</th>
<th>Average</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>W0</td>
<td>W2</td>
</tr>
<tr>
<td>R0</td>
<td>18.00±12.73</td>
<td>38.00±16.97</td>
</tr>
<tr>
<td>R1</td>
<td>10.50±2.12</td>
<td>40.50±36.06</td>
</tr>
<tr>
<td>R2</td>
<td>5.50±2.12</td>
<td>31.00±29.70</td>
</tr>
<tr>
<td>R3</td>
<td>0.25±0.21</td>
<td>5.00±0.10</td>
</tr>
<tr>
<td>R4</td>
<td>0.15±0.07</td>
<td>11.50±6.36</td>
</tr>
<tr>
<td>average</td>
<td>17.28a</td>
<td>19.67a</td>
</tr>
</tbody>
</table>

abc Different superscript at the same raw and column indicate significantly different (P<0.05) according to LSD.
Table 1 shows the addition of artificial preservatives significantly affect the number of fungal colonies during storage. The addition of turmeric and garlic phytobiotic lower than the control and the addition of calcium propionate. Control treatment there is a tendency number of fungal colonies increased more rapidly with increasing storage time. Storage time significantly affect the number of fungal colonies. The addition of garlic lower 26.06 CFUx10³/g significantly different compared with the control and the addition of calcium propionate. This shows that the addition of preservatives affect the rate of mold growth during storage. Storage long relationship with the number of fungal colonies in feed is shown by the equation Y = 1.771x³-11.74x² + 25.56x + 6.88 with a correlation of R² = 0.705. During storage up to 6 weeks of the growth rate of the number of fungal colonies move slowly with the addition of turmeric and garlic phytobiotic, but should still consider the activity of the active substances contained components.

Turmeric has antifungal activity to inhibit the growth of fungi (Wasilah et al., 2007). The greater the concentration of turmeric in the medium, the amount of active substance which diffuses into fungal cells increased resulting in fungal cells become hypertonic and various mechanisms of interference occur in yeast cells, which causes disruption of mold growth and even cause death.

Fungal colony growth in line with the amount of time there was a trend for an increase in water content, temperature and humidity. the level of contamination by fungi are largely determined by the temperature of storage, water and oxygen (Suparjo, 2008).

Alisin antimicrobial activity in their pure form, among others, as an anti-bacterial, antifungal, antiparasitic and antiviral. The antibacterial alisin reported to be effective against a large number of Gram positive and Gram negative including Salmonella, Staphylococcus, Streptococcus, Klebsiela, Proteus, Bacillus, Clostridium, Coliform and Lactobacillus, as well as some anaerobic bacteria (Ankri and Mirelman, 1999; Bongiorno, 2008; Saravanan et al., 2010).

Action antimicrobial garlic is mainly caused by a chemical reaction between sulfur components alisin with groups of sulfur (thiol) of enzymes in microbes, such as trypsin and proteases that will affect the activity of cysteine proteinase involved virulence of some microbes (Ankri and Mirelman, 1999; Davis, 2005), which in turn causes the inhibition of microbial growth (Bakri and Douglas, 2005).

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REFERENCES


